

# Accumulation of Lignans in Suspension Cultures of *Linum mucronatum* ssp. *armenum* (Bordz.) Davis

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For the first time callus and suspension cultures of *Linum mucronatum* ssp. *armenum* were initiated, grown in darkness at 25 °C and analyzed for lignans. 6-Methoxypodophyllotoxin was the main lignan besides smaller amounts of podophyllotoxin isolated and identified by chromatographic methods as well as by <sup>1</sup>H NMR.

## Introduction

Podophyllotoxin (PTOX) is needed as a precursor for chemical synthesis of the important anticancer drugs etoposide, teniposide and etopophos<sup>R</sup> (Canel *et al.*, 2000). Podophyllotoxin is still extracted from rhizomes and roots of *Podophyllum hexandrum* and *P. peltatum* plants collected in the wild, because agricultural production of the plant material as well as chemical synthesis of PTOX are not economic. Therefore, there is a great interest in alternative sources of PTOX supply. Here we report about the initiation of suspension cultures from the Turkish plant species *Linum mucronatum* ssp. *armenum*, their growth characteristics and the accumulation of 6-methoxypodophyllotoxin (MPTOX) as the main lignan besides smaller amounts of podophyllotoxin (PTOX) (Fig. 1).

## Results and Discussion

Sterile grown seedlings of *L. mucronatum* ssp. *armenum* were used for initiation of tissue cul-

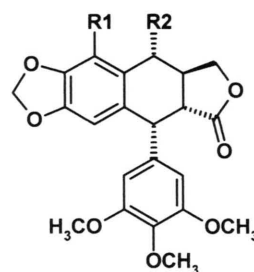


Fig. 1. Podophyllotoxin (R1 = H; R2 = OH), 6-methoxypodophyllotoxin (R1 = OCH<sub>3</sub>; R2 = OH).

tures. In contrast to experience with other *Linum* species (Smolny *et al.*, 1998; Konuklugil *et al.*, 1999) an organogenic callus with leaflets developed, which did not lose its organogenic capacity completely, even not under longer subcultivation in suspension. Therefore, under the experimental conditions used these *in vitro* cultures may be referred to as shooty suspension cultures.

Two lignans were isolated from extracts of these suspension cultures using TLC and HPLC. They were identified by retention time and *r<sub>f</sub>* in HPLC and TLC, respectively (see Experimental), as well as by <sup>1</sup>H NMR (Table I) using reference compounds and in comparison with published data (e.g. Wichers *et al.*, 1991) as 6-methoxypodophyllotoxin (main product) and podophyllotoxin (minor compound). Nomenclature and numbering of both lignans is according to the UPAC recommendations 2000 (Moss, 2000).

Table 1. <sup>1</sup>H-NMR data.

**6-Methoxypodophyllotoxin:** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 6.44 (2H, s, H-2' + H-6'), 6.30 (1H, s, H-3), 5.95 (2H, s, CH<sub>2</sub>(pip)), 5.03 (1H, d, *J* = 8.8 Hz, H-7), 4.64 (1H, dd, *J* = 6.9, 8.8 Hz, H-9a), 4.54 (1H, d, *J* = 4.4 Hz, H-7'), 4.16 (3H, s, OCH<sub>3</sub> at C-6), 4.07 (1H, dd, *J* = 8.8, 10.1 Hz, H-9b), 3.81 (3H, s, OCH<sub>3</sub> at C-4'), 3.77 (6H, s, OCH<sub>3</sub> at C-3' and C-5'), 2.89 (1H, m, H-8), 2.75 (1H, dd, *J* = 4.4, 15.1 Hz, H-8').

**Podophyllotoxin:** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 7.11 (1H, s, H-6), 6.52 (1H, s, H-3), 6.37 (2H, s, H-2' + H-6'), 5.99 (1H, d, *J* = 1.3 Hz, H-a (pip)), 5.97 (1H, d, *J* = 1.3 Hz, H-b (pip)), 4.78 (1H, d, *J* = 8.8 Hz, H-7), 4.62 (1H, dd, *J* = 6.9, 8.8 Hz, H-9a), 4.60 (1H, d, *J* = 4.4 Hz, H-7'), 4.10 (1H, dd, *J* = 8.8, 10.1 Hz, H-9b), 3.81 (3H, s, OCH<sub>3</sub> at C-4'), 3.76 (6H, s, OCH<sub>3</sub> at C-3' and C-5'), 2.84 (1H, dd, *J* = 4.4, 15.1, H-8'), 2.78 (1H, m, H-8).



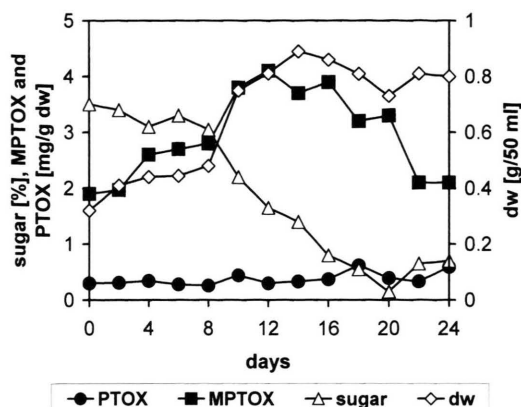


Fig. 2. Growth curve (dry weight) of dark grown suspension cultures of *Linum mucronatum* ssp. *armenum* and the amount of podophyllotoxin (PTOX) and 6-methoxy-podophyllotoxin (MPTOX) of the cells and the sugar content of the medium (%).

The suspension cultures reached about 0.9 g dry weight per 50 ml culture volume after 14 days of culture. The sugar of the medium was taken up by the cells until days 18/20. The cells contained about 4.1 mg/g dw MPTOX on day 12, whereas PTOX content was between about 0.3 and 0.6 mg/g dw throughout the culture period (Fig. 2). The total yield of lignans is comparable to those reported earlier for suspension cultures of *Linum album* and *L. nodiflorum*, respectively (Smolny *et al.*, 1998; Konuklugil *et al.*, 1999).

## Experimental

Callus and suspension cultures were established using standard methods (Seitz *et al.*, 1985) from seeds of *Linum mucronatum* ssp. *armenum* (Bordz.) Davis collected near Ankara and germinated under sterile conditions in 1997. Initiation of callus and suspension cultures as well as isolation and identification of lignans by  $^1\text{H}$  NMR was as reported earlier (Konuklugil *et al.*, 1999). HPLC was performed as described by Smolny *et al.* (1992), the  $R_t$  for PTOX was about 8.2, for MPTOX about 10.3 min under the experimental conditions used. Additionally, for isolation of lignans TLC on silica gel plates (Merck TLC-plates no. 5715) and the solvent systems chloroform:methanol (10:1, v:v;  $R_f$ -value for MPTOX = 0.93 for PTOX = 0.70), chloroform:n-hexane (10:1, v:v;  $R_f$  of MPTOX = 0.29; PTOX = 0.11), and diethylether:dichloromethane (6:1, v:v;  $R_f$  of MPTOX = 0.87; PTOX = 0.69) were used.

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